

Intramolecular H-bonding in the bay region of hypericin: an AM1 study

Dao-Ping Wang^a, De-Xin Kong^a, De-Zhan Chen^a, Hong-Yu Zhang^{b,c,*}

^aDepartment of Chemistry, Shandong Teachers' University, Jinan 250014, PR China

^bCollege of Life Sciences, Shandong Teachers' University, Jinan 250014, PR China

^cLaboratory for Computational Biology, Shandong Provincial Research Center for Bioinformatic Engineering and Technique, Zibo University, Zibo 255091, PR China

Received 21 November 2000; received in revised form 4 January 2001; accepted 24 May 2001

Abstract

The semi-empirical quantum chemical method AM1 was employed to study intramolecular H-bonding in the bay region of hypericin (HYP). It was found that three isomers of HYP, which correspond to structures with 0, 1, and 2 H-bonds in the bay region, readily interconvert, and that the isomer with two H-bonds is the predominant species in polar or nonpolar solvents. It was also shown that the formation of intramolecular H-bonding in the bay region facilitates proton dissociation and intramolecular proton transfer, providing an alternative explanation for the experimentally observed transient species produced from HYP © 2001 Elsevier Science Ltd. All rights reserved.

Keywords: Hypericin; Intramolecular hydrogen bond; Intramolecular proton transfer; Perylenequinonoid pigments; pKa value

1. Introduction

Perylenequinonoid pigments, including hypocrellin A, hypocrellin B, cercosporin, hypericin (HYP) and isohypericin (iso-HYP) (Fig. 1) are photosensitizers that have attracted considerable attention in recent years [1–7]. Structure–activity relationship studies indicate that 4,9-dihydroxy-3,10-perylenequinone is the active center of perylenequinonoid pigments [7–9], and that fast intramolecular proton transfer in the ground state of perylenequinonoid pigments facilitates

intersystem crossing ($S_1 \rightarrow T_1$), enhancing the quantum yield of the T_1 state of such pigments [7]. In addition, the acidity of the hydroxyl group of perylenequinonoid pigments increases in the excited state, which is a beneficial feature in photodynamic therapy [7,10,11].

Although the various perylenequinonoid pigments are similar in structure, they exhibit distinct photosensitization characteristics. For example, the energy barrier to intramolecular proton transfer involving hypocrellin in the excited state is 2.12 ± 0.070 kcal/mol, while that for HYP is only 0.044 ± 0.008 kcal/mol [12]. The pKa for HYP is approximately 6 units lower than that of hypocrellin A, hypocrellin B, and cercosporin [13,14]. Since chemical and physical properties are normally related to molecular structures, it is likely

* Corresponding author. Tel.: +86-533-2780271; fax: +86-533-2780271.

E-mail address: zhysdtu@jn-public.sd.cninfo.net (H.-Y. Zhang).

that the characteristics of HYP arise from its unique structure, in that HYP has two OH groups in the bay region (positions 1 and 2, Fig. 1). It has been found that the facile ionization of HYP is due to the relative proximity of the OH groups, one of which is H-bonded to the O atom of the second OH in a sterically crowded environment [13]. Results of calculations of theoretical heats of formation for intramolecular H-bond formation in the bay region suggest that HYP structures possessing intramolecular H-bonds in the bay region are especially stable [15].

Despite the aforementioned studies, several questions regarding the intramolecular proton transfer process remain unanswered. For instance: (1) What is the energy barrier for intramolecular H-bond formation? (2) Is it possible for HYP to form two intramolecular H-bonds in the bay region (cf. Fig. 2)? (3) How does intramolecular H-bonding influence the photophysics of HYP, including proton dissociation and intramolecular proton transfer? AM1 calculations have been used to address these questions, the results of which are summarized in this paper. Since HYP isomers

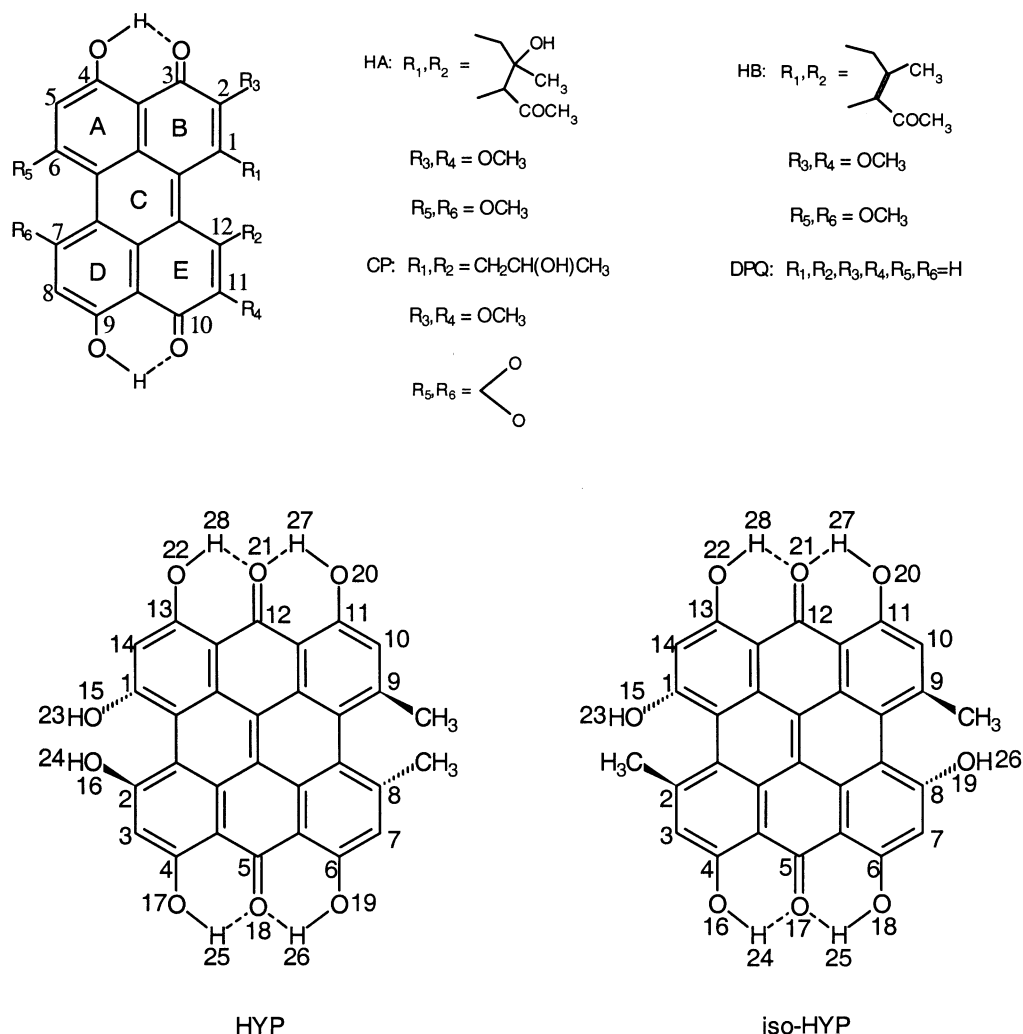


Fig. 1. The structures of perylenequinonoid pigments, including hypocrellin A (HA), hypocrellin B (HB), cercosporin (CP), dihydroxy-perylenequinone (DPQ), HYP, and iso-HYP.



2. Methods

3. Results and discussion

The hydroxyl groups in the bay region of HYP may exist as indicated in Fig. 2, where HYP0, HYP1 and HYP2 arise from 0, 1, and 2 H-bonds in the bay region, respectively. For the inter-conversion of these isomers, ΔG_1 and ΔG_2 are the changes in Gibbs free energy and k_1 and k_2 are the corresponding equilibrium coefficients. The results summarized in Table 1 indicate that enthalpy (H) and entropy (S) decrease as the number of H-

	H (kJ/mol)	S (J/mol)	G (kJ/mol)
HYP0	264.78	747.07	42.04
HYP1	263.63	741.94	42.42
HYP2	260.58	740.31	39.86

Table 3 contains the dipole moments for three HYP isomers. The data show that increasing the number of H-bonds enhances the dipole moment. Since polar solvents stabilize structures that have

large dipole moments, the relative proportions of the 3 isomers would change with solvent polarity.

It is believed that the Gibbs free energy of HYP in a homogeneous medium of dielectric constant ϵ_r can be compared with the Gibbs free energy in a similar medium ($\epsilon_r = 1$) using Eq. (1) [19]:

$$\Delta G_{\text{solv}} = -(N_0/4\pi\epsilon_0) \times (\mu^2/r^3) \times [(\epsilon_r - 1)/(2\epsilon_r + 1)] \quad (1)$$

where ΔG_{solv} is the Gibbs free energy change in a polar solvent, μ is the dipole moment, r is the molecular radius, ϵ_0 is the vacuum permittivity, and N_0 is Avogadro's number. In this study, methanol was used as the solvent, $\epsilon_r = 32.66$, and $r = 6 \text{ \AA}$ [20]. The results of these calculations are shown in Table 3, where G , ΔG_{solv} and G_{solv} correspond to the Gibbs free energy without solvent effects, the change in Gibbs free energy, and the Gibbs free energy with the solvent effects, respectively. We found that G_{solv} decreases as the number of H-bonds increases. In this case, $\Delta G_1 = 0.28 \text{ kJ/mol}$, $\Delta G_2 = -2.65 \text{ kJ/mol}$, $k_1 = 0.89 \text{ s}^{-1}$, $k_2 = 2.91 \text{ s}^{-1}$ and, the proportions of HYP0, HYP1 and HYP2 are 22.32, 19.87 and 57.81%, respectively, which is a

little different from the proportion of these isomers in the absence of solvent effects.

3.2. Influence of intramolecular H-bonding on proton dissociation

It is interesting to note that the pKa value for HYP (~ 1.7) is much lower than that of iso-HYP (7.5) [13], which possesses only one OH group in the bay region, and hypocrellin A (8.22), which has no OH group in the bay region [14]. Thus, it is evident that intramolecular H-bonding in the bay region plays an important role in lowering the pKa of HYP.

The AM1 calculated dissociation free energies for the three isomers of HYP and iso-HYP are given in Table 4. It is clear that the dissociation free energies of HYP1 and HYP2 are much lower than those associated with HYP0 and iso-HYP. This is consistent with the experimental results, which show that intramolecular H-bonding is beneficial to proton dissociation. The dissociation free energy for HYP1 is a little lower than that for HYP2 because the dissociation of the HYP2 proton is inhibited by intramolecular H-bonding. The dissociation free energy for HYP0 is even higher than that for iso-HYP, suggesting that the acidity of HYP0 is weaker than that for iso-HYP.

Having established that the three isomers of HYP coexist in solution, we turned our attention to a study of their monoanions. The free energies for the monoanions (HYP0[−] and HYP1[−]) are listed in Table 4. These values indicate that HYP1[−] is more stable than HYP0[−] by 26.08 kJ/mol. Thus, the equilibrium constant for converting HYP0[−] to

Table 2

Energy barriers for intramolecular H-bond formation in the bay region of HYP (kJ/mol)

HYP	HYP0→HYP1	HYP1→HYP2
Ground state ^a	189.11	188.81
Transition state ^a	204.02	196.31
Energy barrier	14.91	7.50

^a Includes ZPE correction.

Table 3

Gibbs free energy for the three isomers of HYP in methanol

	Dipole moment (debye)	G (kJ/mol) ^a	ΔG_{solv} (kJ/mol) ^b	G_{solv} (kJ/mol) ^c
HYP0	0.32	42.04	−0.014	42.03
HYP1	0.89	42.42	−0.11	42.31
HYP2	1.23	39.86	−0.20	39.66

^a Without solvent effects.

^b With solvent effects.

^c Total free energy in solution.

Table 4

Dissociation free energy for protons in the bay region of HYP and iso-HYP (kJ/mol)^a

	Parent molecule	Anion	Dissociation free energy	pKa [13]
HYP0	42.04	−195.92	1053.39	
HYP1	42.42	−222.00	1026.93	
HYP2	39.86	−222.00	1029.49	1.7
iso-HYP	46.72	−204.04	1041.59	7.5

^a Gibbs free energy for H⁺ is 1291.35 kJ/mol, at $T = 298.15 \text{ K}$.

HYP1[−] is $3.55 \times 10^4 \text{ s}^{-1}$, making HYP1[−] the predominant monoanion in solution.

We also calculated the dissociation free energies for the top protons (H(25) and H(28)) of HYP. In the case of H(28) (Table 5), the proton dissociation free energy involving HYP2 is lower than that of the corresponding protons in HYP0, HYP1 and iso-HYP. However, in each case, the free energy is much higher than that involving the protons in the bay region, suggesting that the top protons are more difficult to dissociate than protons in the bay region. In fact, hypocrellin A has no hydroxyl group in the bay region, and its pKa is much higher than that of HYP and iso-HYP.

3.3. The influence of intramolecular H-bonding on proton transfer

Intramolecular proton transfer involving dihydroxy-perylenequinones is a key factor in the photosensitivity of perylenequinonoid pigments [15]. Until now, only the proton transfer involving

the top protons (H(28)) has been studied [12,21–37]. Among the unresolved questions is whether the experimentally observed transient species arises from intramolecular proton transfer [38]. From the structure of HYP1[−], it can be assumed that intramolecular proton transfer involving a proton in the bay region of HYP1[−] can also occur. Therefore, we compared the intramolecular proton transfer processes involving the H(28) and bay region protons.

The AM1 calculated energy barriers for intramolecular proton transfer involving the H28 protons are 94.13 and 81.69 kJ/mol for HYP2 and HYP1[−], respectively (Table 6). However, the energy barrier for the proton in bay region of HYP1[−] is only 28.56 kJ/mol (Table 6), making it much lower than that for the H28 protons. The rather low energy barrier for HYP1[−] arises from the fact that the O(15)–O(16) distance (2.362 Å) is much shorter than that between oxygens O(16) and O(15) (>2.494 Å) [13]. Although the results of AM1 calculations have only qualitative significance, they suggest intramolecular proton transfer in the bay region is an alternative explanation for the observed transient species produced from HYP. In addition, the X-ray structure of HYP1[−] indicates that the O(15)–H(24) and H(24)–O(16) distances are 1.18 and 1.21 Å, respectively. These values are very similar to those from the AM1 calculated intramolecular proton transfer transition state involving HYP1[−], which are 1.21 and 1.23 Å. This supports the hypothesis that the proton in the bay region of HYP1[−] can undergo intramolecular proton transfer.

Table 5

Dissociation free energy for H(28) of HYP and iso-HYP (kJ/mol)^a

	Parent molecule	Anion	Dissociation free energy
HYP0	42.04	−162.96	1086.35
HYP1	42.42	−159.10	1089.83
HYP2	39.86	−178.91	1072.58
iso-HYP	46.72	−158.48	1086.15

^a The Gibbs free energy of H⁺ is 1291.35 kJ/mol, $T = 298.15 \text{ K}$.

Table 6

Energy barriers (kJ/mol) to intramolecular proton transfer in HYP2 and HYP1[−]

	HYP2 ^a	HYP1 ^{−b}	HYP1 ^{−c}
Ground state	−899.25	−75.32	−75.32
Transition state	−805.12	6.37	−46.76
Energy barrier	94.13	81.69	28.56

^a H(28) transfer from O(22) to O(21) in HYP2, including ZPE correction.

^b H(28) transfer from O(22) to O(21) in HYP1, including ZPE correction.

^c H(24) transfer from O(16) to O(15) in HYP1[−], including ZPE correction.

Acknowledgements

This research was supported by the Natural Science Foundation of the Shandong Province through Grant Nos. Q98D01133 and Q99B06.

References

- [1] Duran N, Song PS. Hypericin and its photodynamic action. *Photochem Photobiol* 1986;43:677–80.
- [2] Jiang LJ. Structure, character, photochemical reaction

- and reaction mechanisms of hypocrellin (I)—structure and character of hypocrellin. *Chin Sci Bull* 1990;35:1608–16 (in Chinese).
- [3] Jiang LJ. Structure, character, photochemical reaction and reaction mechanisms of hypocrellin (II)—reaction of hypocrellin. *Chin Sci Bull* 1990;35:1681–90 (in Chinese).
- [4] Diwu ZJ, Lown JW. Hypocrellins and their uses in photosensitization. *Photochem Photobiol* 1990;52:609–16.
- [5] Diwu ZJ, Lown JW. Phototherapeutic potential of alternative photosensitizers to porphyrins. *Pharmac Ther* 1994;63:1–35.
- [6] Diwu ZJ. Novel therapeutic and diagnostic applications of hypocrellins and hypericins. *Photochem Photobiol* 1995; 61:529–39.
- [7] Zhang HY, Zhang ZY. The structure–activity relationship and broad prospects in applications of photosensitization of perylenequinonoid derivatives. *Adv Free Radic Life Sci* 1999;7:41–7 (in Chinese).
- [8] Diwu ZJ, Lown JW. Photosensitization with anticancer agents 15. Perylene-quinonoid pigments as potential photodynamic therapeutic agents: formation of semi-quinone radicals and reactive oxygen species on illumination. *J Photochem Photobiol B: Biol* 1993;18:131–43.
- [9] Zhang HY. Quantum chemistry calculation study on photosensitization of perylenequinonoid derivatives. *Acta Biochim Biophys Sin* 1998;30:272–6 (in Chinese).
- [10] Sureau F, Miskovsky PS, Chinsky L, Turpin PY. Hypericin-induced cell photosensitization involves an intracellular pH decrease. *J Am Chem Soc* 1996;118:9484–7.
- [11] Chaloupka R, Sureau F, Kocisova E, Petrich JW. Hypocrellin A photosensitization involves an intracellular pH decrease in 3T3 cells. *Photochem Photobiol* 1998;68:44–50.
- [12] Das K, Ashby KD, Wen J, Petrich JW. Temperature dependence of the excited-state intramolecular proton transfer reaction in hypericin and hypocrellin A. *J Phys Chem B* 1999;103:1581–5.
- [13] Freeman D, Frolow F, Kapinus E, Lavie D, Lavie G, Meruelo D, Mazur Y. Acidic properties of hypericin and its octahydroxy analogue in the ground and excited states. *J Chem Soc Chem Commun* 1994:891–2.
- [14] Zhang HY, Zhang ZY. Determination of pK_a values of perylenequinonoid photosensitizer in excited state. *Chin Sci Bull* 1997;42:2005–9.
- [15] Zhang HY. Theoretical study on structure change of hypericin. *Acta Chim Sin* 1999;57:667–71.
- [16] Gajewski JJ, Gilbert KE, McKelvey J. MMX. an enhanced version of MM2 *Adv Mol Model* 1990;2:65–92.
- [17] Dewar MJS, Zoebisch EG, Healy EF, Stewart JJP. AM1: a new general purpose quantum mechanical molecular model. *J Am Chem Soc* 1985;107:3902–9.
- [18] Frisch MJ, Trucks GW, Schlegel HB, Gill PMW, Johnson BG, Robb MA, et al. Gaussian 94, revision E.3. Pittsburgh, PA: Gaussian, Inc, 1995.
- [19] Reichardt C. Solvents and solvent effects in organic chemistry. Weinheim: VCH, 1988. p. 96.
- [20] Reichardt C. Solvents and solvent effects in organic chemistry. Weinheim: VCH, 1988. p. 408–10.
- [21] Gai F, Fehr MJ, Petrich JW. Ultrafast excited-state processes in the antiviral agent hypericin. *J Am Chem Soc* 1993;115:3384–5.
- [22] Gai F, Fehr Mi, Petrich JW. Observation of excited-state tautomerization in the antiviral agent hypericin and identification of its fluorescence species *J Phys Chem* 1994; 98:5784–95.
- [23] Gai F, Fehr Mi, Petrich JW. Role of solvent in excited-state proton transfer in hypericin *J Phys Chem* 1994;98: 8352–8.
- [24] Kraus GA, Zhang W, Fehr Mi, Petrich JW, Wanne-muehler Y, Carpenter S. Research at the interface between chemistry and virology: development of a molecular flashlight *Chem Rev* 1996;96:523–35.
- [25] Das K, English DS, Fehr MJ, Smirnov AV, Petrich JW. Excited-state processes in polycyclic quinones: the light-induced antiviral agent, hypocrellin, and a comparison with hypericin. *J Phys Chem* 1996;100:18275–81.
- [26] Das K, English DS, Petrich JW. Solvent dependence on the intramolecular excited-state proton or hydrogen atom transfer in hypocrellin. *J Am Chem Soc* 1997;119:2763–4.
- [27] English DS, Zhang W, Kraus GA, Petrich JW. Excited-state photophysics of hypericin and its hexamethoxy analog: intramolecular proton transfer as a nonradiative process in hypericin. *J Am Chem Soc* 1997;119:2980–6.
- [28] English DS, Das K, Zenner JM, Zhang W, Kraus GA, Larock RC, Petrich JW. Hypericin, hypocrellin, and model compounds: primary photoprocesses of light-induced antiviral agents. *J Phys Chem A* 1997;101:3235–40.
- [29] Das K, English DS, Petrich JW. Deuterium isotope effect on the excited-state photophysics of hypocrellin: evidence for proton or hydrogen atom transfer. *J Phys Chem A* 1997;101:3241–5.
- [30] English DS, Das K, Ashby KD, Park J, Petrich JW, Castner EW. Conformation of excited-state proton transfer and ground-state heterogeneity in hypericin by fluorescence upconversion. *J Am Chem Soc* 1997;119: 11585–90.
- [31] Das K, Dertz E, Paterson J, Zhang W, Kraus GA, Petrich JW. Hypericin, hypocrellin, and model compounds: steady-state and time-resolved fluorescence anisotropies. *J Phys Chem B* 1998;102:1479–84.
- [32] Petrich JW, Gordon MS, Cagle M. Structure and energetics of ground-state hypericin: comparison of experiment and theory. *J Phys Chem A* 1998;102:1647–51.
- [33] Das K, Smirnov AV, Snyder MD, Petrich JW. Picosecond linear dichroism and absorption anisotropy of hypocrellin: toward a unified picture of the photophysics of hypericin and hypocrellin. *J Phys Chem B* 1998;102:6098–106.
- [34] Zhang HY, Zhang ZY. Theoretical study of intramolecular proton transfer of perylenequinone. *Sci Chin (series B)* 1998;41:85–90.
- [35] Zhang HY. Changes of dipole moment in intramolecular proton transfer process of perylenequinone. *Chin Sci Bull* 1998;43:1380–4.

- [36] Zhang HY. Variance of charges in the process of isomerization of erylenequinone. *J Photochem Photobiol A: Chem* 1999;126:27–30.
- [37] Liu W, Zhang HY, Chen DZ, Zhang ZY, Zhang MH. Theoretical study on intramolecular hydrogen transfer of NH₂-substituted perylenequinone. *Dyes and Pigments* 2000;47:277–84.
- [38] Zhang HY, Chen DZ. Mechanism for intramolecular proton transfer involving hypericin. *Dyes and Pigments* 2000;46:17–21.